Cooperative Interaction of trp Melastatin Channel Transient Receptor Potential (TRPM2) With Its Splice Variant TRPM2 Short Variant Is Essential for Endothelial Cell Apoptosis

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Rationale: Oxidants generated by activated endothelial cells are known to induce apoptosis, a pathogenic feature of vascular injury and inflammation from multiple pathogeneses. The melastatin-family transient receptor potential 2 (TRPM2) channel is an oxidant-sensitive Ca\(^{2+}\)-permeable channel implicated in mediating apoptosis; however, the mechanisms of gating of the supranormal Ca\(^{2+}\) influx required for initiating of apoptosis are not understood.

Objective: Here, we addressed the role of TRPM2 and its interaction with the short splice variant TRPM2 short variant (TRPM2-S) in mediating the Ca\(^{2+}\) entry burst required for induction of endothelial cell apoptosis.

Methods and Results: We observed that TRPM2-S was basally associated with TRPM2 in the endothelial plasmalemma, and this interaction functioned to suppress TRPM2-dependent Ca\(^{2+}\) gating constitutively. Reactive oxygen species production in endothelial cells or directly applying reactive oxygen species induced protein kinase C-\(\alpha\) activation and phosphorylation of TRPM2 at Ser 39. This in turn stimulated a large entry of Ca\(^{2+}\) and activated the apoptosis pathway. A similar TRPM2-dependent endothelial apoptosis mechanism was seen in intact vessels. The protein kinase C-\(\alpha\)-activated phosphoswitch opened the TRPM2 channel to allow large Ca\(^{2+}\) influx by releasing TRPM2-S inhibition of TRPM2, which in turn activated caspase-3 and cleaved the caspase substrate poly(ADP-ribose) polymerase.

Conclusions: Here, we describe a fundamental mechanism by which activation of the trp superfamily TRPM2 channel induces apoptosis of endothelial cells. The signaling mechanism involves reactive oxygen species--induced protein kinase C-\(\alpha\) activation resulting in phosphorylation of TRPM2-S that allows enhanced TRPM2-mediated gating of Ca\(^{2+}\) and activation of the apoptosis program. Strategies aimed at preventing the uncoupling of TRPM2-S from TRPM2 and subsequent Ca\(^{2+}\) gating during oxidative stress may mitigate endothelial apoptosis and its consequences in mediating vascular injury and inflammation. (Circ Res. 2014;114:469-479.)

Key Words: apoptosis ■ capillary permeability ■ endothelium ■ inflammation

Melastatin-like transient receptor potential 2 (TRPM2) is an oxidant-sensitive Ca\(^{2+}\)-permeable channel expressed in many cells, including neurons,1,2 microglia,3,4 multiple lung cell types,5,6 pancreas \(\beta\) cells,7,9 hematopoietic and immune cells,10,11 and vascular endothelial (VE) cells.5 However, the function of TRPM2 remains enigmatic. TRPM2 is activated by the generation of reactive oxygen species (ROS), such as H\(_2\)O\(_2\) and production of adenosine diphosphate ribose (ADPR) after DNA damage and activation of the enzyme poly(ADPR) polymerase.6,12 TRPM2 has been implicated in mediating of oxidant-induced apoptosis secondary to Ca\(^{2+}\) influx that may initiate apoptosis program via the caspase pathway.1,13,14 Although apoptosis is important in normal biological processes and development, apoptosis of endothelial cells, which have low turnover in vessels,1,15 is a fundamental pathogenic feature of inflammatory and vascular diseases, such as acute lung injury16 and sepsis.17 Our studies have demonstrated a key role of TRPM2 in mediating oxidative injury of the endothelium,5 resulting in disruption of endothelial barrier and tissue edema.18-20 A component of endothelial disruption seen in these studies may well have been because of TRPM2-induced apoptosis.

TRPM2 channel opening after exposure to H\(_2\)O\(_2\) and other ROS is induced by the binding of ADPR to the Nudix box sequence motif (nucleoside diphosphate type motif 9 protein) in the carboxyl-terminal domain of TRPM2.5,6,10,12,21-23

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H₂O₂, produced in the cell, also activated the production of ADPR, which functioned by binding to the TRPM2 Nudix motif. In addition, other mechanisms of TRPM2 activation, such as direct oxidative modification of the channel, have been proposed.

Besides TRPM2, several splice variants of TRPM2 associated with TRPM2 in the plasma membrane have also been identified. Their role in regulating TRPM2 function and mediating oxidant-induced apoptosis remains obscure. Of particular interest is the short splice variant (TRPM2-S) that functions as a dominant-negative to inhibit TRPM2 channel activity but which itself lacks both the carboxyl terminus present in the long isoform TRPM2 and the Ca²⁺-permeable pore present in TRPM2. In cells in which both isoforms are expressed, TRPM2-S interact with TRPM2 to inhibit formation of functional homotetrameric channels. Here, we investigated the interaction of TRPM2-S with TRPM2 and how the component cooperated to signal oxidant-induced apoptosis in endothelial cells. The study presents a new mechanism of endothelial apoptosis involving ROS-induced protein kinase C (PKC)-α phosphorylation-dependent disruption of the interaction of TRPM2 with TRPM2-S and opening of the channel to allow sufficient Ca²⁺ entry required for activation of the apoptosis program.

Methods
An expanded Materials and Methods is available in the online Data Supplement.

Endothelial Cell Culture and Transfection
Isolation of Mouse Endothelial Cells
Endothelial cells were isolated from lungs of wild-type (WT), PKCα−/− (obtained from Dr Jeffrey D. Molkentin, University of Cincinnati, Cincinnati, OH) and TRPM2−/− mice (GlaxoSmithKline). The cells were used between passages 2 and 5.

Transfections
Human pulmonary artery endothelial cells (HPAEC; Clonetics, La Jolla, CA) were cultured in gelatin-coated flasks and used between passages 3 and 6. Human TRPM2-S splice variant, tagged with poly-His (His⁶-TRPM2-S), was inserted into a pcDNA3 expression vector (Invitrogen). Phosphorylation-defective TRPM2-S was generated by alanine substitution (S39A), and phosphorylation-mimetic TRPM2-S was generated by aspartic substitution (S39D). Transfection of TRPM2-S constructs using FuGENE HD was verified by Western blotting. Control cells received vector alone.

siRNA Experiments
HPAECs were transiently transfected with TRPM2 or PKCα siRNAs (100 nmol/L; Santa Cruz Biotechnology, Santa Cruz, CA) using TransIT-TKO transfection reagent (Mirus, Madison, WI); nontarget-specific siRNA served as control (Ambion, Austin, TX). Transfection efficiency was >75%.

Immunoprecipitation and Phosphorylation Studies
Untransfected, His⁶-(S39A)TRPM2-S and His⁶-(S39D)TRPM2-S–transfected HPAEC cultures were treated with 300 µmol/L H₂O₂ for indicated times (37°C). In some experiments, cells were pretreated with 3.4-dihydro-S-[4-(1-piperidinyl)butoxy]-1(2H)-isoquinoline or PKC inhibitors 30 minutes before the assay. In other experiments, cells first received siRNA to suppress TRPM2 or PKCα expression. TRPM2 or PKCα immune complexes were precipitated with protein A-Sepharose beads (Sigma) for 2 hours at 4°C as described.

Generation of H₂O₂ Using Glucose Oxidase
H₂O₂ production in vitro was induced by glucose (1 mmol/L) and glucose oxidase (GO; 1–2.5 mU/mL) and was measured spectrophotometrically from the generation of resorufin (absorbance, 565 nm; extinction coefficient, 58,000 mol/L per cm). GO produced H₂O₂ at a constant rate (320 nmol/L/min).

Analysis for Apoptosis
Apoptosis was identified by double-fluorescent staining with phycoerythrin annexin V-fluorescein isothiocyanate and 7-aminoactinomycin D, which detected apoptotic and dead cells, respectively. Confluent endothelial monolayers, without or with PKCα inhibition or silencing, were incubated in 300 µmol/L H₂O₂ for 6 or 24 hours (37°C). Cells were washed twice with PBS and trypsinized; samples of 1×10⁶ cells were incubated with 5 µL of phycoerythrin-labeled annexin V and 5 µL of 7-aminoactinomycin D (BD Bioscience, Rockville, MD) for 20 minutes at 24°C in the dark and analyzed with a Beckman Coulter Cyan II cytometer (Beckman Coulter, Miami, FL). We also assessed apoptosis in intact lung vascular endothelium by immunofluorescence and terminal deoxynucleotidyl transferase dUTP nick-end labeling assay. Lungs of TRPM2−/− and WT mice were perfused (2 mL/min; 37°C) for 3 hours with recirculation of Roswell Park Memorial Institute medium 1640 (5 mL) containing H₂O₂ (300 µmol/L) or GO. Lungs were removed and frozen by the optimal cutting temperature method. Frozen lungs, sectioned (5 µm), were permeabilized with 0.1% Triton X-100. Tissue sections were blocked with 10% fetal bovine serum and incubated with goat anti–VE-cadherin and rabbit anti–poly(ADP-ribose) polymerase (PARP; ie, the cleaved 89-kDa fragment; 1:200 dilution) overnight (4°C). The sections were incubated with secondary antibodies conjugated to Alexa Fluor 488 and 594 (Invitrogen). As an alternative to cleaved PARP antibody, terminal deoxynucleotidyl transferase dUTP nick-end labeling staining was performed according to the manufacturer’s protocol (Roche Diagnostics Corp, Indianapolis, IN). Nuclei were visualized by 4,6-diamidino-2-phenylindole (Sigma-Aldrich, Saint Louis, MO). Slides were analyzed under a Zeiss fluorescence microscope using AxioVision software. Apoptotic cells were identified by double staining (PARP+VE-cadherin or terminal deoxynucleotidyl transferase dUTP nick-end labeling+VE-cadherin).

Statistical Analysis
Statistical comparisons were made with the 2-tailed Student t test. The significance level was P<0.05.

Institutional Study Approval
All studies were conducted after review by the Institutional Animal Care and Use Committee at the University of Illinois at Chicago where the work was performed and in accordance with the Policy on the Care, Welfare, and Treatment of Laboratory Animals.

Methods in Supplement
Methods for [Ca²⁺] measurements, TRPM2 protein purification, in vitro phosphorylation assay, bone marrow transplantation, murine...
model of endotoxin-mediated mortality, and Western blotting are given in the Online Data Supplement.

Results

TRPM2 Is Required for H$_2$O$_2$-Induced Endothelial Cell Apoptosis

Apoptosis was determined by staining endothelial cells with annexin V-phycocerythrin and 7-aminoactinomycin D followed by fluorescence-activated cell sorting assessment (Figure 1A and 1B). H$_2$O$_2$ in a concentration-dependent manner induced apoptosis within 24 hours with an EC$_{50}$ value of 136±6 μmol/L (Figure 1A). Inhibition of TRPM2 by an anti-TRPM2 blocking antibody or TRPM2 siRNA silencing prevented the apoptosis (Figure 1B). The flow cytometry dot plot data demonstrating apoptosis are shown in Online Figure I. The sustained generation of H$_2$O$_2$ (320 nmol/L per minute for 90 minutes) by GO with glucose substrate also induced endothelial apoptosis, which was blocked by TRPM2 silencing or inhibition of channel activity (Figure 1B). To address in vivo relevance, we also examined apoptosis in endothelial cells of lung vessels in WT and TRPM2 knockout mice perfused with H$_2$O$_2$ or GO/glucose infusion in physiological stimulus to generated oxidants. Here, we used tumor necrosis factor-α (TNF-α), a generator of intracellular oxidants and potent inducer of endothelial cell apoptosis. We determined expression of the 17- and 20-kDa caspase-3 fragment and cleaved PARP in lungs of WT and not TRPM2 –/– mice (Figure 1D). Using another apoptosis assay, terminal deoxyribonucleotidyl transferase dUTP nick-end labeling staining, we observed fewer endothelial cells undergoing apoptosis after H$_2$O$_2$ or GO/glucose infusion in TRPM2 –/– mice when compared with WT (Online Figure II).

H$_2$O$_2$ Induces PKCα Phosphorylation of TRPM2-S

To address whether H$_2$O$_2$ was involved in the phosphorylation at Ser39 on TRPM2-S, we used HPAEC monolayers treated with PKCα inhibitors (Gö6976 or PKCα blocking peptide [PKCα inhibitor peptide]) or transfected with siRNA to knockdown PKCα expression. PKCα was immunoprecipitated from lysates of cells exposed to H$_2$O$_2$ and coimmunoprecipitated TRPM2 and TRPM2-S were detected using an antibody recognizing each form (Figure 2B). H$_2$O$_2$ rapidly induced the association of PKCα with TRPM2-S, but not with TRPM2, and the response persisted up to 5 minutes (Figure 2B). Immunoprecipitation was reduced when PKCα activation was inhibited (Figure 2B). Treatment with PCKβIII inhibitory peptide, used as control for nonspecific effects of Gö6976 in blocking activation of both PKCα and PCKβIII, did not modify the H$_2$O$_2$-induced TRPM2-S association with PKCα (Figure 2B). PKCα-TRPM2-S immunoprecipitation was also suppressed predictably by TRPM2 silencing (Figure 2B). Control experiments showed that transfection of endothelial cells with TRPM2 siRNA significantly reduced the expression of both TRPM2 and TRPM2-S (Figure 2A). Control experiments confirmed that siRNA effectively suppressed PKCα expression (Figure 2B). Treatment of cells with PKCα inhibitors (Gö6976 or PKCα inhibitor peptide) and PCKβIII blocking peptide did not modify PKCα expression (Figure 2B).

Because the above studies dealt with the role of H$_2$O$_2$ in activating the phosphorylation of TRPM2-S, we next examined whether key alterations could be replicated using a physiological stimulus to generated oxidants. Here, we used tumor necrosis factor-α (TNF-α), a generator of intracellular oxidants and potent inducer of endothelial cell apoptosis. We observed that TNF-α induced the association of PKCα with TRPM2-S, whereas suppressing PKCα activity prevented the association (Online Figure IIIA).

To determine whether PKCα was responsible for phosphorylating TRPM2-S after H$_2$O$_2$ challenge, in other studies TRPM2 proteins in cell lysates were precipitated using antibodies recognizing TRPM2 and TRPM2-S, and phosphorylated proteins were visualized using antiphospho-Ser antibody. Western blotting demonstrated that only TRPM2-S was phosphorylated, which occurred within 1 minute of H$_2$O$_2$ exposure with maximum response seen at 2 minutes, whereas there was no phosphorylation of TRPM2 (Figure 2C). Western blotting also showed that TRPM2-S but not TRPM2 was phosphorylated within the same time frame after TNFα exposure (Online Figure IIIIB). Phosphorylation of PKCα (82 kDa) was detected as a comigrating band on the gel (seen in top blot of Figure 2C; Online Figure IIIIB), an indication that the kinase was in the active state.

We next determined phosphorylation of PKCα using an antibody recognizing phosphorylated on Ser 657, the crucial PKCα catalytic domain. H$_2$O$_2$ rapidly induced phosphorylation of PKCα at this site and the phosphorylated PKCα co-migrated with TRPM2 (middle blot; Figure 2C). Treatment with Gö6976 (but not with control PCKβIII inhibitor peptide) inhibited not only PKCα phosphorylation but also H$_2$O$_2$-induced phosphorylation of TRPM2-S (Figure 2C). These results thus show time-dependent and reversible association between TRPM2-S and PKCα induced by PKCα activation (Figure 2D).
S39 Phosphorylation of TRPM2-S Activates TRPM2 and Supranormal Ca²⁺ Influx

We used the Fura-2 dye to study the Ca²⁺ entry response activated by TRPM2 interaction with TRPM2-S. In addition, we used the Ca²⁺ add-back protocol to rule out any indirect effects of H₂O₂ on Ca²⁺ entry secondary to Ca²⁺-store depletion. In the absence of extracellular Ca²⁺, H₂O₂ did not produce a Ca²⁺ transient (Figure 3A and 3B), indicating that H₂O₂ did not deplete intracellular Ca²⁺ stores. By contrast, extracellular Ca²⁺ repletion in the continued presence of H₂O₂ elicited a sharp and marked increase in intracellular Ca²⁺ concentration secondary to Ca²⁺ entry (Figure 3A and 3B). TRPM2 knockdown markedly suppressed the Ca²⁺ transient (Figure 3A and 3B), showing that H₂O₂-induced Ca²⁺ entry required TRPM2. G66976 significantly decreased the amplitude of Ca²⁺-repletion-dependent transients by 66±9%, and PKCα inhibitor peptide reduced Ca²⁺ transient by 46±10% (Figure 3A and 3B). PKCα silencing also reduced Ca²⁺ entry...
PKCα Regulated TRPM2 Gating and Apoptosis

Treatment of cells with control PKCβII peptide inhibitor, however, did not modify H2O2-activated Ca2+ entry via TRPM2 channels (Figure 3A and 3B). Along the same lines, TNFα-induced Ca2+ entry in endothelial cells (Online Figure IID) was also decreased by inhibiting PKCα.

We next addressed the role of the S39 phosphoswitch on TRPM2-S in mediating TRPM2 channel activity. Here, we determined whether mutation of TRPM2-S at Ser 39 (S39A), the PKCα phosphorylation site disrupted Ca2+ signaling. The mutant was tagged on its C terminus with a poly-His fusion protein. Transfected HPAECs showed protein expression of the (S39A)-TRPM2-S mutant (Figure 4A). Western blotting showed that S39A mutation of TRPM2-S abrogated the migration of TRPM2-S with PKCα on the gel (Figure 4B) and phosphorylation of TRPM2-S by PKCα after H2O2 challenge (Figure 4C). PKCα also did not migrate with (S39A)-TRPM2-S mutant (Figure 4C).
with the TRPM2-S phosphodefective mutant (Figure 4D).

To validate the finding that PKCα was indeed responsible for phosphorylation of TRPM2-S at S39, we treated an extract of native protein with recombinant active PKCα. We observed that active PKCα induced phosphorylation of WT TRPM2-S but not of S39A mutant (Online Figure IV).
PKCα Phosphorylation of TRPM2-S Induces TRPM2-S Dissociation From TRPM2

We next determined whether PKCα phosphorylation of TRPM2-S in some manner interfered with TRPM2-S association with TRPM2, thus permitting TRPM2 to gate Ca\(^{2+}\) at sufficient level to activate the apoptosis program. Because TRPM2 variants generated by alternative splicing differed only in their C terminal,\(^28\) we immunoprecipitated TRPM2 from cell lysates using an anti-TRPM2 antibody, recognizing the region present solely in TRPM2 form. TRPM2-S, which in the plasma membrane basally associated with TRPM2, dissociated within minutes from TRPM2 after H\(_2\)O\(_2\) addition (Figure 5A). Inhibition of PKCα activation suppressed this TRPM2-S dissociation from TRPM2 (Figure 5A). S39A mutation of TRPM2-S also suppressed the dissociation of TRPM2 from TRPM2-S (Figure 5A). PKCα-dependent phosphorylation of TRPM2-S at Ser 39 blocked the interaction of TRPM2-S with TRPM2 (summarized in Figure 5B). These results show that phosphorylation of TRPM2-S at Ser 39 was responsible for releasing the TRPM2-S inhibition of TRPM2 and thus mediated the increased Ca\(^{2+}\) entry needed for apoptosis.

To reinforce the crucial role of TRPM2-S S39 phosphorylation in mediating TRPM2 channel activity, we mutated TRPM2-S S39 to aspartate (S39D) to mimic the effects of phosphorylation. This poly-His tagged phosphomimetic mutant was expressed in HPAECs (Online Figure V4A), and we examined its ability to associate with TRPM2, and influence Ca\(^{2+}\) entry. Western blotting showed that phosphomimetic phosphorylation mutants with either PKCα siRNA or control siRNA in endothelial cells (Online Figure VI). Expression of (S39A)-TRPM2-S phosphodefective mutant in control siRNA-transduced cells as expected inhibited H\(_2\)O\(_2\)-elicited Ca\(^{2+}\) entry when compared with control cells, whereas expression of phosphomimetic mutant enhanced this response. The decreased H\(_2\)O\(_2\)-activated Ca\(^{2+}\) entry caused by depletion of PKCα was restored by expression of the (S39D)-TRPM2-S but not the (S39A)-TRPM2-S mutant, consistent with the essential role of PKCα phosphorylation of TRPM2-S in activating TRPM2 channel activity.

PKCα Mediates H\(_2\)O\(_2\)-Induced Apoptosis Through Activation of TRPM2

Fluorescence-activated cell sorting analysis showed that inhibition of PKCα activation or its silencing protected the cells from H\(_2\)O\(_2\)-induced apoptosis (Figure 6A and 6B). The role of PKCα in regulating TRPM2-mediated apoptosis was also seen in endothelial cells transduced with the (S39A)-TRPM2-S mutant (Figure 6A). Using lung endothelial cells cultured from TRPM2 or PKCα knockout mice to validate the above studies in human endothelial cells, we observed normal expression of TRPM2 in endothelial cells from PKCα knockout mice and normal expression of PKCα in endothelial cells from TRPM2 knockout mice (Figure 6A and 6B). The H\(_2\)O\(_2\)-induced Ca\(^{2+}\) entry was virtually abolished in TRPM2-null cells and was reduced by 40% in PKCα-null cells (Figure 6C). As in the human cells, H\(_2\)O\(_2\)-mediated apoptosis in WT mouse endothelial cells was concentration dependent (Figure 6D). Deletion of TRPM2 caused 2.4-fold rightward shift in the concentration-response curve for H\(_2\)O\(_2\)-induced apoptosis (EC\(_{50}\) shift, 213–553 µmol/L), indicating the crucial role of TRPM2 in mediating H\(_2\)O\(_2\)-induced apoptosis (Figure 6D). Deletion of the PKCα gene similarly inhibited apoptosis (EC\(_{50}\) shift, 213–512 µmol/L; Figure 6D).

Deletion of PKCα Gene in Mice Reduces TRPM2-Induced Endothelial Cell Apoptosis Improves Survival in Endotoxemia

To address the pathophysiological significance of PKCα phosphorylation of TRPM2 channel activity in mediating apoptosis in vivo, we examined the apoptosis response in mouse lung endothelial cells and survival of mice after intraperitoneal challenge with lipopolysaccharide (30 mg/kg), the Gram-negative bacterial endotoxin, which produces ROS in endothelial cells.\(^34,35\) Because TRPM2 and PKCα expressed in myeloid cells may also play a role in ROS production and apoptosis,\(^36\) we generated chimeric mice in which the PKCα- and TRPM2-deficient mice were transplanted with bone marrow cells from WT mice. These mice showed comparable TRPM2 and PKCα protein expression as WT (Figure 7A). We observed that either TRPM2 or PKCα...
deletion markedly reduced endothelial cell apoptosis in lungs 4 hours after lipopolysaccharide treatment when compared with WT mice (Figure 7B). In a positive control experiment, administration of the oxidant scavenger Tempol in mice 30 minutes before lipopolysaccharide also reduced oxidant-mediated lipopolysaccharide-induced apoptosis (Figure 7B). In addition, deletion of either TRPM2 or PKCα significantly improved survival rate of lipopolysaccharide-challenged mice (Figure 7C).

Discussion
In the present study, we addressed the role of the ROS-activated TRPM2 channel in mediating endothelial cell apoptosis. We identified that the interaction of the 171-kDa TRPM210,12,27,37 with its 90-kDa splice variant TRPM2-S28 in the endothelial cell plasma membrane. This interaction functioned constitutively to restrain TRPM2 Ca2+ entry. However, ROS-induced activation of PKCα and resulting phosphorylation of TRPM2-S at Ser39 released the TRPM2-S inhibition of TRPM2 to induce the large Ca2+ influx required for activation of the caspase apoptosis program.

PKCα phosphorylation of TRPM2-S and the dissociation of TRPM2-S from TRPM2 increased the Ca2+ concentration in endothelial cells to 4-fold the baseline levels within the range of the Ca2+ burst required to signal apoptosis, which has the intracellular Ca2+ concentration threshold of 200 to 500 nmol/L.38 Inhibition of PKCα by preventing the phosphorylation of TRPM2-S reduced Ca2+ entry through TRPM2 by half this level, well below the Ca2+ threshold required for activation of caspase-mediated apoptosis.

A stop codon (TAG) on the TRPM2 gene is located at the splice junction between exons 16 and 17; hence, alternative splicing resulted in deletion of the 4 C-terminal transmembrane domains in TRPM2-S, the putative Ca2+-permeable pore region.28 The observation that TRPM2-S served as a dominant-negative for TRPM2, and its uncoupling from TRPM2 required for the full gating of Ca2+ identifies TRPM2-S as an important intrinsic negative regulator of endothelial cell apoptosis. H2O2 exposure or oxidants generated by mediators, such as TNF-α, induced the interaction between PKCα and TRPM2-S permitting the channel to open for Ca2+ entry. Thus, TRPM2-S only functioned to induce apoptosis when PKCα was activated and induced TRPM2-S phosphorylation. That TRPM2-S mediated Ca2+ gating through heterodimerization with TRPM2 is reminiscent of the finding in melanocytes that another splice variant member of the trp gene family TRPM1 interacted with full-length long form of TRPM1 to suppressed its activity.39

PKCα activation was shown to be crucial for the mechanism of H2O2-induced apoptosis through its binding to and phosphorylation of TRPM2-S. Mutation of the sole PKCα phosphorylation site on Ser39 of TRPM2-S N terminus to

Figure 6. H2O2-induced endothelial cell apoptosis resulting from melastatin-like transient receptor potential 2 (TRPM2)-mediated Ca2+ influx. Human pulmonary artery endothelial cells (HPAECs; A) and mouse lung endothelial cells (B–D) challenged with H2O2 were labeled with phycoerythrin (PE)-annexin/7-aminoactinomycin D (7-AAD). A, Flow-cytometry histograms (left) and summary plots of apoptosis (right) 0, 6, and 24 hours after challenge with H2O2 (300 µmol/L) or glucose oxidase/glucose to generate 320 nmol/L H2O2/min for 90 minutes, as a function of PKCα inhibition or silencing (±SEM; n=5). B–D, Mouse endothelial cells were isolated from lungs of TRPM2−/−, PKCα−/−, and wild-type (WT) mice. B, Western blot verifying absence of PKCα expression in PKCα−/− cells and of TRPM2 expression in TRPM2−/− cells. C, Left, Ca2+ mobilization assay using Ca2+ add-back protocol with the fluor-3 Ca2+ indicator. Right, Mean ratiometric values (±SEM) for steady-state [Ca2+]i (n=6). *P<0.0001 vs WT cells (t test). D, Dose–response curve for H2O2-induced apoptosis in mouse endothelial cells detected by flow cytometry. Deletion of PKCα or TRPM2 caused ~2.5-fold rightward shift in the dose–response curve.
Ala resulted in failure of PKC\(\alpha\) to phosphorylate TRPM2-S. This mutation in turn prevented PKC\(\alpha\)-mediated apoptosis. Both K562 myeloid leukemia cell line and Jurkat T-lymphocyte cell line that expresses the short isoform at very low levels\(^{14,40}\) also did not undergo apoptosis secondary to TRPM2 activation,\(^{14,40}\) consistent with the critical role of TRPM2-S as the apoptosis-suppressing partner of TRPM2. Although PKC\(\alpha\) activation contributed to TRPM2-induced endothelial apoptosis in the present study, it is important to note that PKC\(\alpha\) signaling in other signaling pathways can induce endothelial injury. We have shown that PKC\(\alpha\) phosphorylation of p120-catenin mediates disassociation of p120-catenin from VE-cadherin that resulted in disassembly of adherens junctions and resultant increased lung permeability.\(^{41}\) Thus, PKC\(\alpha\) can function in a complex manner at multiple levels to induce endothelial dysfunction either via injury or through activating the apoptosis program.

The signaling pathway downstream of Ca\(^{2+}\) entry leading to cell death involves the activation of intrinsic executioner caspases (caspase-9) and extrinsic caspases (caspase-8) that activate the effector caspase-3 and caspase-7.\(^{14,42-44}\) These cleave cellular substrates, disrupting survival pathways and inducing membrane

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Figure 7. Protein kinase C-\(\alpha\) (PKC\(\alpha\)) interaction with melastatin-like transient receptor potential 2 (TRPM2) in mice is required lung endothelial cell apoptosis in response to lipopolysaccharide (LPS) and contributes to mortality. WT, TRPM2\(^{-/-}\), and PKC\(\alpha\)-mice were transplanted with bone marrow cells isolated from WT mice 8 weeks before Western blotting (A), apoptosis (B and C), and survival (D) studies. A, Representative Western blots verifying expression of TRPM2, PKC\(\alpha\), and \(\beta\)-actin in bone marrow of transplanted mice (molecular masses of 171, 82, and 45 kDa, respectively). Left, Protein was quantified by densitometry. TRPM2 and PKC\(\alpha\) densities were normalized to \(\beta\)-actin and plotted as percentage of untreated control (mean\(\pm\)SEM for \(n=3\)). Endothelial apoptosis (B) and oxidant production (C) were determined in lungs of mice 4 hours after intraperitoneal injection of LPS (40 mg/kg) and in lungs of mice treated with the oxidant scavenger Tempol (100 mg/kg, IP) 30 minutes before injection of LPS. B, Right, Immunofluorescent staining of frozen lung sections using vascular endothelial-cadherin (red) antibody, terminal deoxynucleotidyl transferase dUTP nick-end labeling (green), and 4,6-diamidino-2-phenylindole (DAPI; blue; \(n=3\)). Scale bar, 50 \(\mu\)m. Left, Quantification of apoptotic endothelial cells (mean\(\pm\)SEM; \(n=3\)), *\(P<0.001\) and †\(P<0.005\) vs LPS-treated wild-type (WT) lungs. TRPM2 and PKC\(\alpha\) deletion significantly reduced LPS-induced endothelial apoptosis. C, Lungs were homogenized and assayed for H\(_2\)O\(_2\) accumulation using the horseradish peroxidase-linked Amplex Red assay. H\(_2\)O\(_2\) was determined spectrophotometrically from its absorbance at 570 nmol/L and corrected for total protein (mean\(\pm\)SEM; \(n=3\)). †\(P<0.005\) vs LPS-treated control lungs. D, Deletion of either TRPM2 or PKC\(\alpha\) reduced LPS-induced lethality in mice. LPS (30 mg/kg) was injected intraperitoneally and survival was assessed every 12 hours during the experiment (WT, \(n=16\); TRPM2\(^{-/-}\), \(n=16\); and PKC\(\alpha\)-, \(n=12\)). Statistical analysis was performed using the log-rank test. *\(P<0.03\) and †\(P<0.05\) vs WT cells.
blebbing, cell shrinkage, and apoptotic body formation. PARP is part of a protective mechanism involved in repair of DNA damage⁴⁴ and DNA stability.⁴⁵ Inactivation of PARP by cleavage of the enzymatic domain after oxidant activation of TRPM2 also caused apoptosis¹⁴ similar to that seen with PKC-α-induced uncoupling of TRPM2-S from TRPM2 in the present study.

We have uncovered in these studies a novel mechanism of TRPM2 activation resulting in Ca²⁺ entry secondary to PKC-α-induced phosphorylation of TRPM2-S. A question arises about the relationship of this mechanism with TRPM2 activation induced by the generation of ADPriboselysation after the activation of poly(ADPR) polymerase.⁶¹⁲ It is possible that both mechanisms function to activate TRPM2 secondary ROS stimulation (see Model Figure 8). ADPR generation after activation of poly(ADPR) polymerase may help to amplify the Ca²⁺ entry response. However, in the event that both TRPM2-S and TRPM2 are coexpressed as they are in endothelial cells, it is likely as the present results show that TRPM2-S functions by restraining the activity of TRPM2 (and hence suppresses apoptosis). However, when TRPM2-S is not expressed or poorly expressed, ADPR binding to Nudix box sequence would by default be the primary mechanism of TRPM2 activation, but it is not clear whether Ca²⁺ entry by this mechanism is sufficient to activate the proapoptotic caspases.

In summary, we identified a fundamental relationship between oxidant-activated TRPM2 channel and its associated short splice variant TRPM2-S in the gating of large Ca²⁺ influx and the critical role of loss of this interaction in mediating oxidant-induced apoptosis of endothelial cells. We demonstrated that apoptosis induced by this mechanism contributed to the mortality seen in endotoxin-challenged mice. PKCα functioned to induce phosphorylation of TRPM2-S, which prevented its association with TRPM2, and thereby activated Ca²⁺ gating and caspases. Thus, disabling TRPM2-S and TRPM2 interaction such as by inhibiting PKCα activation represents a novel strategy for abrogating apoptosis and resultant vascular injury and inflammation associated with apoptosis in diseases, such as acute lung injury and vascular inflammation.

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Disclosures

None.

References

phosphorylation of the short splice variant of TRPM2, TRPM2-S, and inflammatory disorders. We have uncovered a role of PKCα in understanding the molecular mechanisms regulating apoptosis and understanding the role of TRPM2 in oxidative stress-induced death.

**Activation of the cation (primarily Ca2+) permeable melastatin transient receptor potential 2 (TRPM2) channel during oxidative stress is linked to cell death.**

**What New Information Does This Article Contribute?**
- Oxidant activation of TRPM2 mediates lung endothelial cell apoptosis and is critically regulated by protein kinase C-α (PKCα).
- TRPM2 in endothelial cells is normally impermeable to Ca2+ because of its binding to the short splice variant TRPM2 short variant (TRPM2-S).
- Oxidants induce PKCα phosphorylation of TRPM2-S at Ser 39, which functions by releasing TRPM2-S inhibition of TRPM2 channel and thereby induces Ca2⁺ entry and sequestration to activate the apoptotic program.

### Novelty and Significance

**What is known?**
- Oxidants induce injury to the vascular endothelium resulting in endothelial denudation, permeability alterations, edema formation, and inflammation.
- Endothelial cell loss via apoptosis is a crucial feature of vascular injury and is implicated in the mechanism of lung injury induced by sepsis and other vascular diseases.
- Activation of the cation (primarily Ca2⁺) permeable melastatin transient receptor potential 2 (TRPM2) channel during oxidative stress is linked to cell death.

**What new information does this article contribute?**
- Oxidant activation of TRPM2 mediates lung endothelial cell apoptosis and is critically regulated by protein kinase C-α (PKCα).
- TRPM2 in endothelial cells is normally impermeable to Ca2⁺ because of its binding to the short splice variant TRPM2 short variant (TRPM2-S).
- Oxidants induce PKCα phosphorylation of TRPM2-S at Ser 39, which functions by releasing TRPM2-S inhibition of TRPM2 channel and thereby induces Ca2⁺ entry and sequestration to activate the apoptotic program.

**Endothelial cell apoptosis is a crucial feature of vascular injury that leads to disruption of the endothelial barrier and to inflammation.** Understanding the molecular mechanisms regulating apoptosis is vital for identifying novel targets for treating vascular injury and inflammatory disorders. We have uncovered a role of PKCα phosphorylation of the short splice variant of TRPM2, TRPM2-S, which acts as a phosphoswitch to regulate channel activity and results in calcium overload. Phosphorylation of TRPM2-S at Ser 39 caused release of TRPM2-S inhibition of TRPM2 and thereby activated Ca2⁺ gating. This unique mechanism of TRPM2 channel activation was crucial for induction of oxidant-mediated apoptosis of endothelial cells. Thus, oxidant-induced gating of Ca2⁺ via TRPM2 and apoptosis are critically dependent on PKCα phosphorylation of TRPM2-S at a specific site identifying a novel potential anti-inflammatory target.
Cooperative Interaction of trp Melastatin Channel Transient Receptor Potential (TRPM2) With Its Splice Variant TRPM2 Short Variant Is Essential for Endothelial Cell Apoptosis
Claudie M. Hecquet, Min Zhang, Manish Mittal, Stephen M. Vogel, Anke Di, Xiaopei Gao, Marcelo G. Bonini and Asrar B. Malik

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SUPPLEMENTAL MATERIAL

Methods

Reagents and chemicals. Endothelial growth medium (EBM-2) was obtained from Clonetics (San Diego, CA), and the Dulbecco's modified Eagle's medium (DMEM) from Invitrogen (Grand Island, NY). His6-tagged TRPM2 cDNA construct was made by modification of the cDNA encoding the green fluorescent protein-fused TRPM2-S (GFP-TRPM2-S) kindly provided by Dr. Barbara A. Miller (Pennsylvania State University College of Medicine, Hershey, PA). Trypsin, Hank’s balanced salt solution (HBSS), molecular cellular and developmental biology (MCDB) media 131, TRizol reagent, AmplexR Red glucose/glucose oxidase assay kit and Superscript II were obtained from Invitrogen (Carlsbad, CA). FuGENE HD transfection reagent and TUNEL assay kit were obtained from Roche Applied Science (Indianapolis, IN); and TransIT-TKO Mirus transfection reagent from Mirus Bio (Madison, WI). Fura-2/acetoxymethyl ester (AM) was obtained from Molecular Probes (Eugene, OR). The myristoylated PKCα peptide inhibitor Myr-RFARKGALRQKNV was from Promega (Madison, WI). H2O2, myristoylated PKCβII inhibitor Myr-SLNPEWNET (PKCβIII), penicillin, lipopolysaccharides (LPS) and peptides and chemicals were from Sigma Chemical Co. (St. Louis, MO). Matrigel, Dynabeads M-450 and the platelet/endothelial cell adhesion molecule-1 (PECAM-1) were purchased from BD Bioscience (San Jose, CA). Anti-TRPM2 antibodies (one against the 171-kDa TRPM2 long isoform and the other recognizing both TRPM2 and TRPM2-S isoforms) were purchased from Abcam (Cambridge, MA). His6, PKCα and phospho-Ser antibodies were from Santa Cruz Biotechnology (Santa Cruz, CA).

Isolation of endothelial cells from mouse lungs. Mice at age 6-8 weeks were deeply anesthetized (2.5% isoflurane in a bell jar), and heparin (50 U/mouse) was injected into the jugular vein. A thoracotomy was carried out and the pulmonary artery was cannulated. Krebs-Henseleit solution supplemented with bovine serum albumin (5 g/100 mL) was infused to remove blood. Lungs were removed and placed inside a culture hood. Lung tissue slices from 3 mice were prepared, washed, and suspended in HBSS. Excess HBSS was aspirated, and the tissue slices were minced and transferred to a 15-mL sterile tube. The minced tissues were suspended in 10 mL of collagenase A (1.0 mg/mL in HBSS) and digested for 60 minutes at 37°C with gentle shaking. The released cells were collected by centrifugation, resuspended, filtered and incubated in growth medium (EGM-2 supplemented with 10% fetal bovine serum), then plated on Matrigel-coated 35-mm culture dish and allowed to confluence for 10 days. Cells were then harvested from the Matrigel plates by dispase (BD Bioscience) for 60 to 90 minutes. Cells were washed after dispase treatment once with growth medium and plated on 0.1% gelatin coated culture dish. Cells passaged between 3 and 4 times were used in experiments. Endothelial cells were characterized by their cobblestone morphology, platelet/endothelial cell adhesion molecule-1 (PECAM-1 or CD31) expression, and Dil-Ac-LDL uptake.

Endothelial cell culture and transfection.

Isolation of endothelial cells from murine tissue: Endothelial cells were isolated from lungs of WT, PKCα−/− (obtained from Dr Jeffrey D. Molkentin 3, University of Cincinnati, Cincinnati, OH) and TRPM2−/− (obtained from GlaxoSmithKline 4) mice as described 5. Cells were cultured in gelatin-coated flasks using DMEM/F12 medium supplemented with endothelial growth factor plus 10% fetal bovine serum, and used in experiments between passages 2–5.

Transfections: Human pulmonary artery endothelial cells (HPAE; Clonetics, La Jolla, CA) were cultured in gelatin-coated flasks using endothelial basal medium 2 (EBM2) supplemented with bullet kit additives plus 10% fetal bovine serum, and used in experiments between passages 3–6. Human TRPM2-S short splice variant, tagged on its carboxy-terminus with poly-His (His6-TRPM2-S), was inserted into pcDNA3 expression vector (Invitrogen). Phosphorylation-defective TRPM2-S was generated by alanine substitution (S39A) and phosphorylation-mimetic TRPM2-S was generated by aspartic substitution (S39D). Point mutation was introduced in His6-TRPM2-S construct using the QuikChange site-directed mutagenesis protocol (Stratagene), and was verified by sequencing. HPAE cell cultures, grown to 60-80% confluency,
were transfected with 1 μg/ml each of His\(^6\)-(S39A)TRPM2-S or His\(^6\)-(S39D)TRPM2-S cDNA, or with vector alone (control cells) using fuGENE HD and in the presence of protease inhibitor cocktail (Sigma Aldrich) to prevent degradation of the transfected protein. In some experiments, cells were co-transfected with PKCa siRNA and TRPM2-S mutant cDNA using X-tremeGENE siRNA Transfection Reagent (Roche) and maintain in culture medium containing protease inhibitor cocktail (Sigma Aldrich) to prevent degradation of the transfected protein and caspase 9 inhibitor (Ac-LEHD-CHO, 20 mol/L) to prevent apoptosis. Successful transfection of cells with (S39A)TRPM2-S or (S39D)TRPM2-S and depletion of PKCa was verified by Western Blot.

**Stable transfection of human HEK293 cells:** HEK293 cells grown at 37°C with 5% CO\(_2\) in DMEM supplemented with 10% fetal bovine serum were transfected with 1 μg/ml each of the human long variant of TRPM2, tagged on its carboxy-terminus with poly-His (His\(^6\)-TRPM2) and inserted into pcDNA6 expression vector (Invitrogen) and either His\(^6\)-TRPM2-S or His\(^6\)-(S39A)TRPM2-S plasmids using the FuGENE HD transfection reagent. The successfully transfected cells were then selected with Geneticin (G418, 100 μg/mL) and Blasticidin (100 μg/mL).

**Small interfering RNA transfection:** HPAEs were transiently transfected with 100 nmol/L of TRPM2 or PKCa pre-designed small interfering RNAs (siRNAs, Santa Cruz Biotechnology, Santa Cruz, CA) using TransIT-TKO transfection reagent (Mirus, Madison, WI) according to manufacturer's instructions. As control, we used commercially available nonspecific (NS) siRNA (Ambion, Austin, TX). Protein silencing was verified by Western Blots analysis. Transfection efficiency was at least 75%.

**[Ca\(^{2+}\)]i measurements.**

**Ratiometric Ca\(^{2+}\) measurements using Fura-2/AM:** Control or transfected HPAE cells (see above) grown to confluence on 25-mm glass coverslips were loaded with Fura-2/AM (2 μmol/L) for 20 min at 37°C. Cells received two washes with Hank’s balanced salt solution and were placed in an experimental chamber containing 200 μl of buffer. We measured Fura 2 fluorescence using Attoflor Ratio Vision digital fluorescence microscope (Atto Instruments, Rockville, MD) equipped with F-Fluar 40 x oil-immersion objectives with a numerical aperture of 1.3. Excitation wavelengths used were 340 and 380 nm, and emission wavelength was 510 nm. Intracellular Ca\(^{2+}\) levels are given as fluorescent ratio F340/F380 representing bound/free Ca\(^{2+}\).

Ca\(^{2+}\) measurements were also made using FlexStation scanning fluorometer. Mouse endothelial cells were grown to confluence in clear-bottom 96-well assay plates. Assays utilized the FLIPR (Fluorometric Imaging Plate Reader) Calcium Plus kit (Molecular Devices, Sunnyvale, CA). Cells were loaded with the FLIPR Ca\(^{2+}\)-sensitive fluorescence indicator and incubated for 1 h at 37°C according to the manufacturer's protocol. The addition of agonists was robotically controlled, and monolayer fluorescence in each well was read by the FlexStation data acquisition system (Molecular Devices) at 0.1 Hz. Cells were excited at 485 nm and monitored at 515 nm.

**Western blotting.** Endothelial monolayers were washed in PBS, lysed in Tris buffer (containing 1% Triton X-100, 1 mmol/L phenylmethylsulfonyl fluoride [PMSF], and protease-inhibitor cocktail), and sonicated (20 s). Protein was separated by electrophoresis (4-12% SDS gradient polyacrylamide gel) and transferred to nitrocellulose membranes for Western blotting with antibodies (TRPM2, PKCa, His\(^6\), phospho-Ser, or actin). Band intensity was determined by densitometry using Image J (NIH).

**Immunoprecipitation and phosphorylation studies.** Untransfected and His\(^6\)-(S39A)TRPM2-S transfected HPAE cultures in six-well culture dishes were treated with 300μM H\(_2\)O\(_2\) for various times at 37°C. In some of the experiments, cells were pretreated with the Poly(ADPR) polymerase inhibitor (DPQ) or PKC inhibitors described above 30 min prior to the assay. In other experiments, cells were previously transduced with siRNA to selectively suppress expression of TRPM2 or PKCa. Following H\(_2\)O\(_2\) challenge, cells were washed with ice-cold PBS and lysed with 0.5% deoxycholate buffer (pH 7.5) containing 1% NP-40, 0.1% SDS, 1 mmol/L PMSF, 50 mmol/L Tris, 150 mmol/L NaCl, and 10 μl protease inhibitor mixture. After shaking for 10 min at 4°C, lysates were sonicated and then centrifuged for 15 min at 16,000 g and 4°C. Supernatants were collected and diluted with 390 μl of 50 mmol/L Tris buffer (pH 7.5) containing 150 mmol/L NaCl and protease inhibitors. Samples were then incubated with 1 μg of antibody (rabbit anti-TRPM2 or anti-PKCa) overnight at 4°C. TRPM2 or PKCa immune complexes were precipitated with protein A-Sepharose beads (Sigma) at 4°C for 2 h. The beads were then washed five times with lysis buffer, and the precipitated proteins were eluted by boiling the beads in sample buffer [80 mmol/L Tris (pH 6.8), 3%
SDS, 15% glycerol, 0.01% bromophenol blue, 5% DTT). Proteins were then separated on a 4–12% SDS-PAGE gradient gel.

**His-Tagged TRPM2 protein purification using Ni-NTA beads:** His<sup>6</sup>-TRPM2-S/ His<sup>6</sup>-TRPM2 transfected HEK cells pelleted from 50 ml tissue culture were resuspended in 8 ml of native binding buffer (50 mM NaH<sub>2</sub>PO<sub>4</sub>, 0.5 M NaCl, pH 8.0), supplemented with protease and phosphatase inhibitors. Cells were lysed by two freeze-thaw cycles. The lysates preparation were passed through an 18-gauge needle to shear the DNA, then centrifuged at 3,000 × g for 15 minutes to pellet the cellular debris. The supernatant (8 ml) was transferred to a 15-mL purification column containing 1 ml of 50% slurry of Ni-NTA beads (Qiagen, Valencia, CA) at 4°C for 2 h. Beads were washed twice with native buffer containing 20 mM imidazole and His<sup>6</sup>-tagged TRPM2 proteins were eluted with native buffer containing 250 mM imidazole. The eluted proteins were stored at −20°C.

**In vitro phosphorylation assay.** For TRPM2 phosphorylation by PKCα, 5μg of the TRPM2 channel proteins (short and long) purified from transfected HEK cells fraction were incubated for 1 h at 30°C in the absence or presence of 0.045 pg of PKCα in a buffer containing 25 mM Hapes (pH 7.4), 1 mmol/L DTT, 10 mmol/L MgCl<sub>2</sub>, 0.2 mmol/L Na<sub>3</sub>VO<sub>4</sub>, 1.7 mmol/L CaCl<sub>2</sub>, 5 mM beta-glycerophosphate. Phosphorylation was initiated by the addition of ATP at a concentration of 50 μmol/L. The reaction was terminated by the addition of sample buffer [0.35 mol/L Tris-HCl (pH 6.8), 10.28% (w/v) SDS, 36% (v/v) glycerol, 0.6 mol/L DTT, 0.012% (w/v) bromophenol blue] and boiling for 5 min. Proteins were separated by SDS-PAGE and analyzed by immunodetection with an anti-phosphoserine antibody after transfer to nitrocellulose.

**Measurement of hydrogen peroxide released.** The amount of H<sub>2</sub>O<sub>2</sub> released after the action of glucose oxidase was measured spectrophotometrically based on the generation of resorufin (absorbance at 565 nm) using the extinction coefficient at 58,000 M<sup>−1</sup>cm<sup>−1</sup> and the height of a 150 μL column of solution in a typical 96-well plate. The concentration of glucose oxidase used in experiments was calculated to produce a continuous flow of 320 nmol/L H<sub>2</sub>O<sub>2</sub>/min in cells.

The amount of H<sub>2</sub>O<sub>2</sub> generated in the mouse lungs 4 h following lipopolysaccharides (LPS, 40 mg/kg) stimulation was measured with a horseradish peroxide-linked Amplex Red assay (Molecular Probes, Carlsbad, CA). Lungs of mice untreated, injected intraperitoneally with LPS and treated with Tempol (100 mg/kg, IP) 30 min before injection of LPS were homogenized in PBS containing protease inhibitors and the Amplex Red dye; H<sub>2</sub>O<sub>2</sub> was determined spectrophotometrically (absorbance at 570 nM) using a microplate reader and corrected for total protein content assessed using the Bradford assay.

**Analysis for apoptotic cell death.** Apoptosis was identified by double fluorescence staining with PE Annexin V-FITC (to detect apoptotic cells) and 7-AAD (to detect dead cells). Apoptotic cells translocate phosphatidylserine from the internal face of the plasma membrane to the outer surface, and therefore stain with Annexin V-PE which binds with high affinity to phosphatidylserine, resulting in red fluorescence when excited at 450–480 nm. Confluent endothelial monolayers in 6-well culture dishes, untreated or treated to inhibit PKCα activation, were exposed to 300 μM H<sub>2</sub>O<sub>2</sub> for 6 or 24 h at 37°C. Following H<sub>2</sub>O<sub>2</sub> challenge, cells were washed twice with PBS and trypsinized; cell samples (1 × 10<sup>6</sup> cells per sample) were incubated with 5 μl of PE-labeled Annexin V and 5 μl of 7-AAD (BD bioscience, Rockville, MD) for 20 min at 24°C in the dark and then analyzed with a Beckman Coulter CyAn II cytometer (Beckman Coulter, Miami, FL) within 1 h of Annexin V-PE labeling.

**Apoptosis in lung endothelium by immunofluorescence and Terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL) assay:** Lungs of TRPM2<sup>−/−</sup> and wild-type mice were perfused (2 ml/min, 37°C) for 3 h with a recirculating volume of RPMI 1640 solution containing H<sub>2</sub>O<sub>2</sub> (300 μmol/L) or glucose oxidase (that produced 320 nmol/L/min H<sub>2</sub>O<sub>2</sub>). Lungs were removed, inflated and frozen using an OCT solution. Frozen lungs were cut (5 μmol/L) and fixed in 3.7% paraformaldehyde, then permeabilized in 0.2% tritonX-100 containing buffer for 5 min. Tissue sections were block in 10% FBS and incubated with the goat anti-VE-cadherin and the rabbit antibody raised against the 89 kDa cleaved-PARP (1:200 dilution) overnight at 4°C. The immunofluorescence assay was performed by incubation with secondary antibodies conjugated to Alexa Fluor 594 and Alexa Fluor 488 (Invitrogen). Nuclei were visualized by 4,6-diamidino-2-phenylindole staining (DAPI, Sigma-Aldrich, Saint Louis, MO). Alternatively to cleaved PARP antibody, TUNEL staining was performed according to the manufacturer’s protocol (Roche Diagnostics Corp.; Indianapolis, IN). Slides were analyzed under Zeiss fluorescence microscope with ApoTome attachment (Axio Imager Z1 stand) and equipped with Axiocam camera and the AxioVision software. Apoptotic cells in the alveolar area were identified by double staining (PARP + VE-cadherin).
Bone marrow transplantation: Bone marrow transplantation was performed as previously described. Recipient mice were lethally irradiated with 9.5 Gy and received an i.v. injection of 4 million donor bone marrow cells (isolated from WT mice) under ketamine / xylazine (100/5 mg/kg IP) anesthesia 24 hr after irradiation. To determine the transplantation efficiency, bone marrow cells were immunoblotted with anti-TRPM2 antibody. LPS injection and survival studies were performed 8 weeks after bone marrow transplantation.

Murine model of endotoxin-mediated mortality: Eight weeks following bone marrow transplantation, mice were challenged with LPS (30 mg/kg body weight) via intraperitonial injection. This LPS concentration (30 mg/kg BW) was established to be lethal. Mice were observed for feeding, movement and activity, grooming (smooth and shiny coats versus dull and ruffled coats) and survival for 96 h.

Statistical analysis. Mean values ± S.E.M were calculated for each experiment and statistical comparisons were made with the two-tailed Student’s t-test. The significance of differences between groups was determined with a two-tailed t-test.

References


Online Figure I: TRPM2 mediates H$_2$O$_2$-induced apoptosis of endothelial cells. **Left**, representative flow cytometry histograms at 0, 6, or 24 h after exposure to 300 µmol/L H$_2$O$_2$ or glucose oxidase/glucose (90 min), with or without prior TRPM2 silencing or inhibition with TRPM2 blocking antibody. **Right panel**, mean percentage of apoptotic cells at 0, 6, or 24 h after H$_2$O$_2$ treatment (± SEM, n=3). The baseline values were not significantly altered by TRPM2 silencing or TRPM2 blocking Ab.
**Online Figure II: Apoptosis in lungs of TRPM2−/− vs. wild-type (WT) mice.** Apoptosis in lungs of TRPM2−/− and wild-type (WT) mice measured 3 h after perfusion with a solution containing H$_2$O$_2$ (300 μmol/L) or glucose oxidase/glucose (75 min). (A) Representative photomicrographs of TUNEL staining in the lung. TUNEL-positive cells are shown in green, VE-cadherin is counterstained in red (Alexa 594) and nuclear in blue (DAPI), (n=3). Scale bar: 50 μm. (B) Percentages of apoptotic endothelial cells; percentages were obtained from 2 fields/slide X 6 (±SEM; n=6). * p = 0.001 compared with control (t-test). Deletion of TRPM2 markedly reduced endothelial apoptosis in lungs of mice challenged with H$_2$O$_2$ or glucose oxidase/glucose compared to WT.
Online Figure III: TNFα induces PKCα phosphorylation of TRPM2-S and TRPM2-dependent Ca²⁺ entry in endothelial cells. HPAECs transduced with PKCα siRNA or untreated with PKC inhibitors (100 nmol/L G60976, 1 μmol/L PKCai) were challenged with 20 ng/mL TNFα for the indicated times at 37°C. (A) TNFα induced the association of PKCα and TRPM2-S. PKCα was immunoprecipitated from cell lysates and co-immunoprecipitated TRPM2 was detected by Western blotting using an antibody recognizing both TRPM2 and TRPM2-S. TRPM2-S associated with PKCα following TNFα exposure whereas inhibition of PKCα prevented the association. (B) TNFα induced PKCα-dependent phosphorylation of 90kDa TRPM2-S splice variant. TRPM2 was immunoprecipitated from same cell lysates using an Ab that recognizes either TRPM2 isoform. (C), Mean densitometric values (± SEM; n=3-4) obtained in A-B showing that PKCα inhibition prevented TNFα-induced association of PKCα with TRPM2-S and phosphorylation of TRPM2-S. (D), Left, Ca²⁺ mobilization assay using “the Fluor-3 Ca²⁺ indicator. Right, Summary of mean ratiometric data (± SEM) for the peak intracellular [Ca²⁺], (n = 6). *P ≤ 0.0005 vs. control (t-test). PKCα inhibition or deletion abrogated TNFα-elicited Ca²⁺ transients.
Online Figure IV: PKCα mediates phosphorylation of TRPM2 at serine 39. His<sup>6</sup>-tagged TRPM2<sup>S</sup> and His<sup>6</sup>-TRPM2<sup>S</sup> (S39A) channel proteins purified from transfected HEK cells were incubated with recombinant PKCα (active, 0.5 μg) in a reaction buffer containing 50 μmol/L ATP for 30 min at 30 °C.

(A) Representative Western blot for PKCα-dependent phosphorylation of these channel proteins analyzed using a specific anti-phospho-serine antibody.

(B) Mean densitometric values of TRPM2<sup>S</sup> and (S39A)TRPM2<sup>S</sup> serine phosphorylation relative to baseline untreated controls (± SEM; n = 3). * p = 0.0001 compared with PKCα treated control (t-test).
Online Figure V. TRPM2-S Ser 39 phospho-mimetic mutant fails to bind TRPM2 and enhances H₂O₂-induced Ca²⁺ entry. The predicted PKCa phosphorylation site on TRPM2-S N-terminus at Ser 39 was mutated by Asp (phospho-mimetic substitution). HPAE monolayers transduced with mutant TRPM2-S (tagged on its carboxy-terminal end with poly-His residues) were grown to confluence and prepared for Western blot analysis (A through C) or intracellular Ca²⁺ measurements using fura-2 (D). (A-C). Cells were exposed to 300 μM H₂O₂ for the indicated times. (A) Western blots for TRPM2, PKCa, and GAPDH expression in cells transduced with phosphomimetic construct. TRPM2 and anti-His⁶ Abs confirmed expression of mutant TRPM2-S construct. (B) TRPM2 was immunoprecipitated from cell lysates using an Ab recognizing both forms of TRPM2 and co-immunoprecipitated PKCa protein was detected with an Ab. Graph in B, mean densitometric values (± SEM; n=3-4). Mutation of Ser 39 with Ala in TRPM2-S prevented TRPM2-S association with PKCa while mutation with Asp increased it. (C) TRPM2 was immunoprecipitated from cell lysates with an anti-TRPM2 Ab recognizing a region present only on the long isoform. The co-immunoprecipitated short isoform was then detected using an Ab that recognizes both TRPM2 and TRPM2-S. Graph in C, density of co-immunoprecipitated TRPM2-S was quantified as ratio to TRPM2 and plotted relative to the zero time value of untransfected control cells (mean ± SEM; n = 3). S39 phosphomimetic mutation of TRPM2-S prevented association of TRPM2-S with TRPM2 at time zero and following H₂O₂ addition. (D) Ca²⁺ mobilization assays were carried out using the “Ca²⁺ add-back” protocol. Transduction of phosphomimetic TRPM2-S mutant showed enhanced H₂O₂-induced Ca²⁺ entry (mean ± SEM; n = 6). *p = 0.0001 compared with control (t-test).
Online Figure VI. Expression of TRPM2-S phospho-mimetic mutant restores H$_2$O$_2$-induced Ca$^{2+}$ entry in PKCα-depleted cells. HPAEC co-transduced with the mutant TRPM2-S and (A) control siRNA or (B) PKCα siRNA were prepared for intracellular Ca$^{2+}$ measurements using “Ca$^{2+}$ add-back” protocol with the Fluor-3 Ca$^{2+}$ indicator. (A) In control cells, transduction of phosphomimetic TRPM2-S mutant showed enhanced H$_2$O$_2$-induced Ca$^{2+}$ entry while transduction of the phoso-defective mutant decreased it (mean ± SEM; n = 6). (B) Depletion of PKCα decreased Ca$^{2+}$ entry elicited by H$_2$O$_2$ compared to control cells; however, the peak Ca$^{2+}$ transient was restored by expression of the (S39D)TRPM2-S but not (S39A)TRPM2-S. (C) Summary of mean ratiometric data (± SEM) for the peak intracellular [Ca$^{2+}$]i obtained in (A-B) (n = 5 to 6). *P ≤ 0.0005 vs. control siRNA (t-test).